

NTM: diagnostic & epidemiology

Samira Boarbi

National Reference Laboratory
Tuberculosis & Mycobacteria

SBIMC-BVIKM SYMPOSIUM 24th May 2022

Diagnostic



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graph TD; Diagnostic --> Species[Species identification]; Diagnostic --> AST[Antibiotic Susceptibility Test (AST)];
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**Species
identification**

**Antibiotic Susceptibility Test
(AST)**

> 200 mycobacterial species

Strictly pathogenic	Potentially pathogenic or opportunist	Non (or rarely) pathogenic	
<p>Slowly growing</p> <p><i>M. tuberculosis</i> <i>M. bovis</i> <i>M. africanum</i> <i>M. ulcerans</i> <i>M. microti</i> <i>M. leprae</i></p> <p><i>M. tuberculosis</i> complex</p>	<p>Slowly growing</p> <p><i>M. asiaticum</i> <i>M. avium</i> <i>M. celatum</i> <i>M. farcinogenes</i> <i>M. genavense</i> <i>M. haemophilum</i> <i>M. interjectum</i> <i>M. intermedium</i> <i>M. intracellulare</i> <i>M. kansasii</i> <i>M. lentiflavum</i> <i>M. lepraemurium</i> <i>M. malmoense</i> <i>M. marinum</i> <i>M. paratuberculosis</i> <i>M. scrofulaceum</i> <i>M. shimoidei</i> <i>M. simiae</i> <i>M. szulgai</i> <i>M. xenopi</i></p> <p>Rapidly growing</p> <p><i>M. abscessus</i> <i>M. chelonae</i> <i>M. fortuitum</i> <i>M. peregrinum</i> <i>M. porcinum</i></p>	<p>Slowly growing</p> <p><i>M. cookii</i> <i>M. gastri</i> <i>M. gordonae</i> <i>M. hiberniae</i> <i>M. nonchromogenicum</i> <i>M. terrae</i> <i>M. triviale</i></p> <p>Rapidly growing</p> <p><i>M. agri</i> <i>M. aichiense</i> <i>M. alvei</i> <i>M. aurum</i> <i>M. austroafricanum</i> <i>M. brumae</i> <i>M. chitae</i> <i>M. chubuense</i> <i>M. confluentis</i> <i>M. diernhoferi</i> <i>M. duvalii</i> <i>M. fallas</i> <i>M. flavescens</i> <i>M. gadium</i> <i>M. gilvum</i> <i>M. komossense</i> <i>M. madagascariense</i> <i>M. methylovorum</i></p>	<p><i>M. morokaense</i> <i>M. neoaurum</i> <i>M. obuense</i> <i>M. parafortuitum</i> <i>M. phlei</i> <i>M. poriferae</i> <i>M. pulveris</i> <i>M. rhodesiae</i> <i>M. shangaiense</i> <i>M. smegmatis</i> <i>M. sphagni</i> <i>M. thermoresistibile</i> <i>M. tokaiense</i> <i>M. vaccae</i> <i>M. yunnanense</i></p>

Non tuberculosis mycobacteria (NTM)

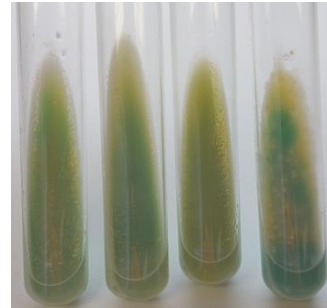
- Slowly growing
- Rapidly growing

Diagnostic



DNA preparation
(Thermolysate)

Sub-culture
on LJ tubes



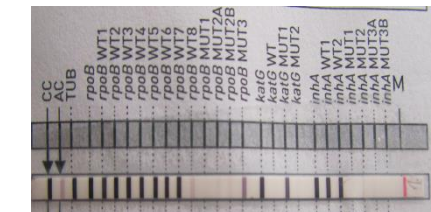
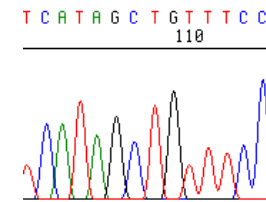
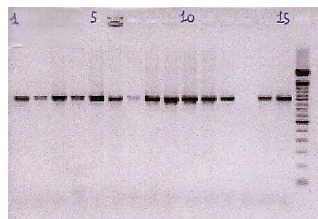
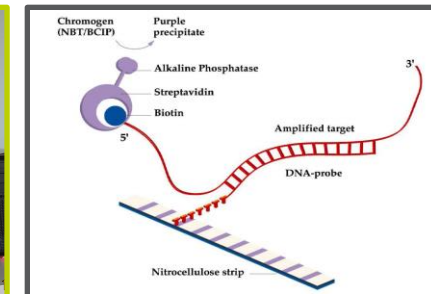
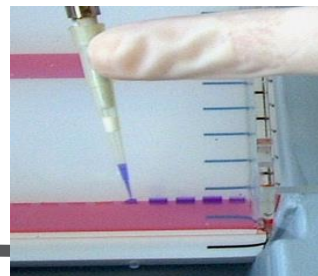
Molecular tests

- ✓ PCR
- ✓ PCR + sequencing
- ✓ PCR + hybridization (LPA)



Species identification

Detection of mutations associated
with the AB resistance



Cultures on solid medium



Different media:

Löwenstein-Jensen (LJ)

Coletsos

Middlebrook 7H10 and 7H11



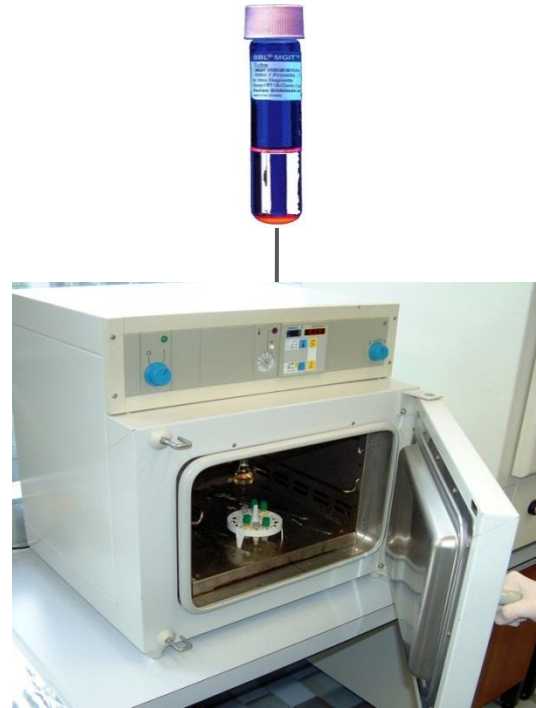
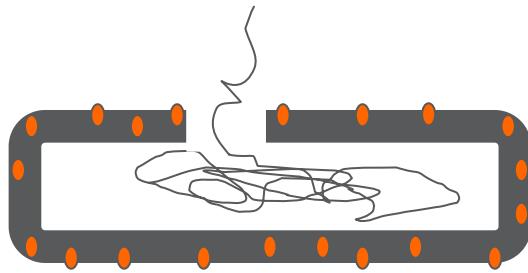
Some mycobacterial species can be distinguished according to the aspect/pigmentation of the culture



Positif : 4-6 weeks

Macroscopic examination possible

Thermolysate : DNA preparation



Culture aliquote at
95° C 30 min

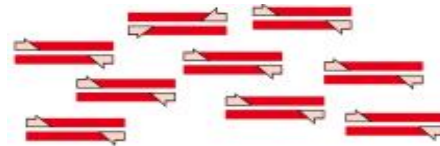
Thermolysate (= inactivated
culture, DNA preparation)

PCR + sequencing

PCR amplification
of region of interest



Purification
Preparation



Sequencing

16SrRNA : species identification

CGG	ATA	GG-ACCA	CGG	GAT	TCA	TG	TCC-	TGT	Séquence déterminée
CGG	ATA	GG-ACCA	CGG	GAT	GCA	TG	TCT-	TGT	<i>M. tuberculosis</i>
CGG	ATA	GG-ACCT	CAA	GAC	GCA	TG	TCT-	TCT	<i>M. avium</i>
CGG	ATA	GG-ACCA	CGG	GAT	TCA	TG	TCC-	TGT	<i>M. marinum</i>
CGG	ATA	GG-ACCT	TTA	GGC	GCA	TG	TCT-	TTA	<i>M. intracellulare</i>
CGG	ATA	GG-ACCA	CTT	GGC	GCA	TG	CCT-	TGT	<i>M. gastri</i>
CGG	ATA	GG-ACCA	CTT	GGC	GCA	TG	CCT-	TGT	<i>M. scrofulaceum</i>
CGG	ATA	GG-ACCA	CTT	GGC	GCA	TG	CCT-	TGT	<i>M. simiae</i>
CGG	ATA	GG-ACCT	CTC	GGC	GCA	TG	CCTA	GGA	<i>M. intermedium</i>

PCR + hybridization = LPA (Line Probe assay)

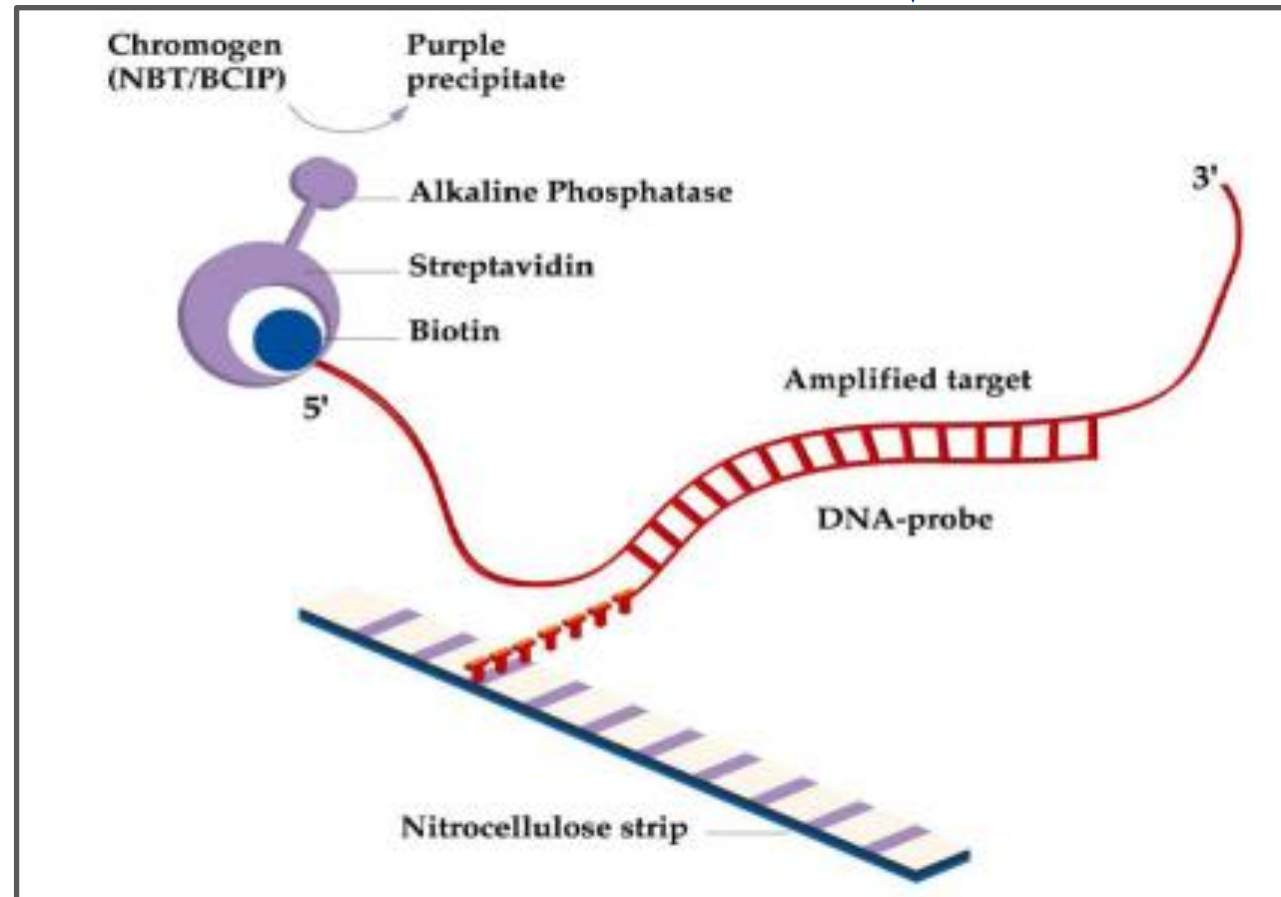
PCR amplification
of region of
interest



Separation of the 2
DNA strands by
heating



Hybridization on
membrane coated
with probes

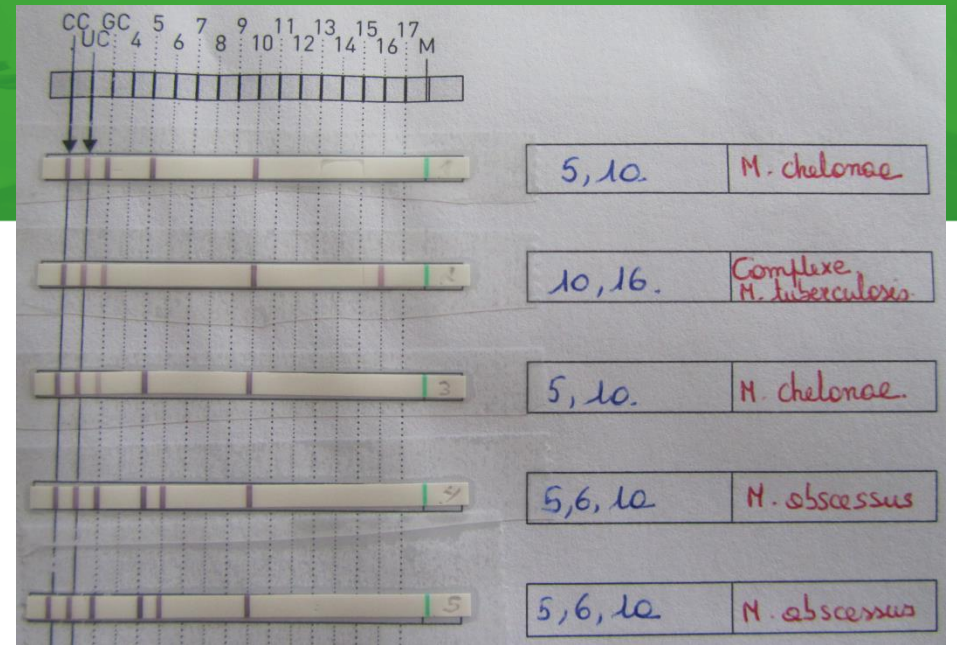


Commercial kits

Species Identification

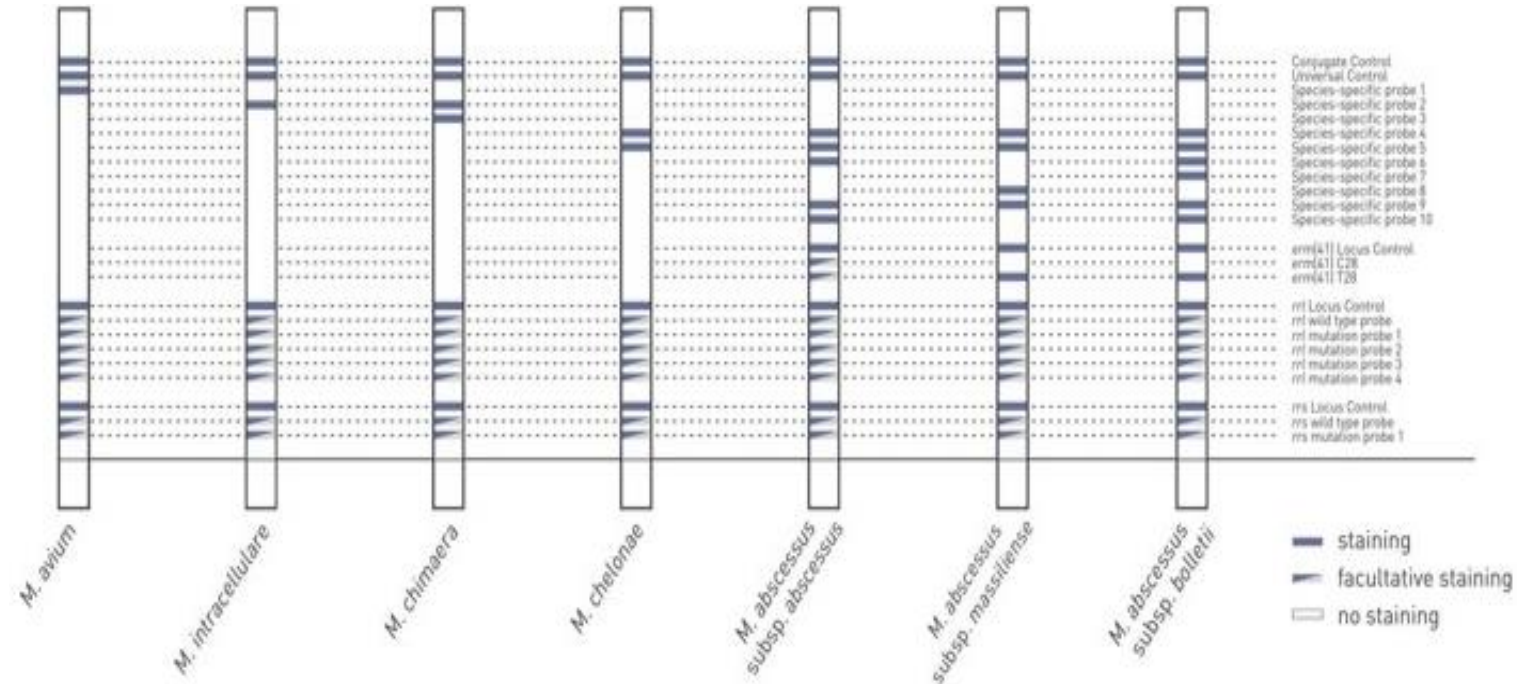
Differentiation of *M. tuberculosis* complex and different NTMs
 - **GenoType Mycobacterium CM/AS (23S RNA gene)**

Differentiation of the *M. avium* and *M. abscessus* complex
 - **GenoType NTM-DR**



Detection of mutations associated with AB resistance

- **GenoType NTM-DR:** resistance to macrolides and aminoglycosides



Commercial kits

- GenoType NTM-DR:

GenoType NTM-DR			
Genes	Probes	Mutations	Resistance
erm(41)	C28	Only Interpretable for <i>M.absc ssp abscessus</i> and <i>bolletii</i> .	Macrolides (clarithro ou azithro)-Sensitive
	T28	<i>M.absc ssp massiliense</i> is deleted in erm(41) so always macrolides-sensitive except if rrl mutation .	Macrolides (clarithro or azithro)-Resistant
rrl	MUT1	A2058C	Macrolides (clarithro or azithro)
	MUT2	A2058G	
	MUT3	A2059C	
	MUT4	A2059G	
	Delta WT	A2058T ou A2059T	
rrs	MUT1	A1408G	Aminoglycosides (kanamycin, amikacine, gentamicin)
	Delta WT	T1406A ou C1409T	

Antibiotic Susceptibility test (AST)

Species identification



Partial report with remark:

The antibiogram will only be performed on request, if clinically relevant.



Opmerkingen / interpretatie:

Een niet-tuberculeuze Mycobacterie geïsoleerd uit een respiratoir staal is niet noodzakelijk de oorzaak van een pathologie. Indien de klinische toestand van de patiënt dit vereist zullen wij op aanvraag een antibiogram uitvoeren.

Antibiotic Susceptibility test (AST)

Proportion method of Canetti (solid medium)

Revue de Tuberculose et de Pneumologie, T. 27, 1963, n° 2-3 (pp. 217-272).

MESURE DE LA SENSIBILITÉ DU BACILLE TUBERCULEUX
AUX DROGUES ANTIBACILLAIRES
PAR LA MÉTHODE DES PROPORTIONS.

MÉTHODOLOGIE, CRITÈRES DE RÉSISTANCE,
RÉSULTATS, INTERPRÉTATION

par

G. CANETTI, N. RIST et J. GROSSET
(Institut Pasteur, Paris).



Broth microdilution method (Sensititre®)



From 15/04/2022, the Canetti technique for the determination of susceptibility to non-tuberculous mycobacteria will no longer be performed due to insufficient scientific evidence for interpretation. The microdilution method (Sensititre®) is recommended by the CLSI. If you have any questions, you can always contact us at BK@sciensano.be

 sciensano

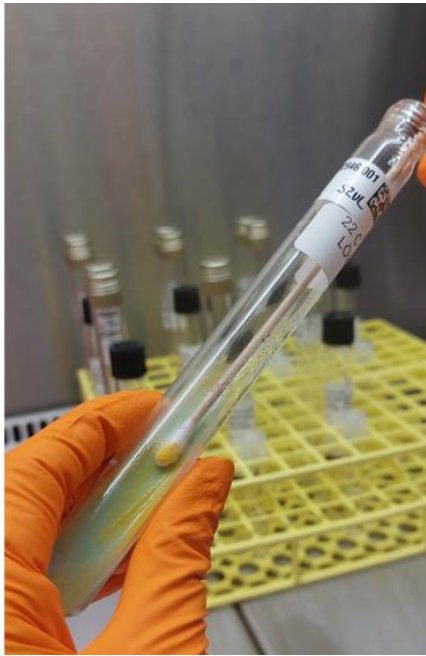


“Broth microdilution is the validated method for AST for NTM”

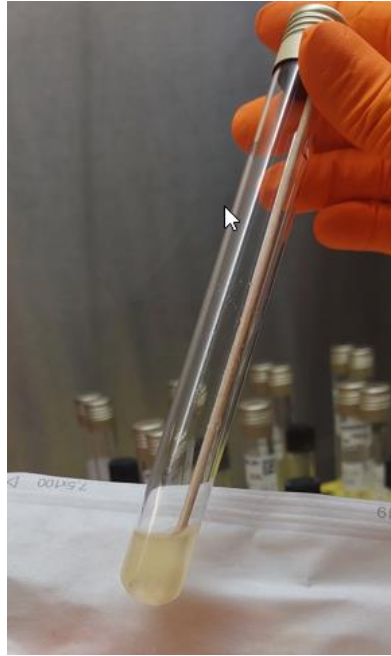
.be

Antibiotic Susceptibility test (AST)

Preparation of the inoculum



Scraping with a swab
the slope of LJ tube



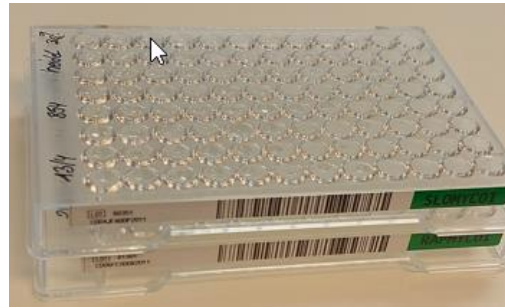
Suspension with glass beads



Adjust to 0.5-1 McFarland



Distribution in the plate



Incubation:

- RGM: 4 to 14 days (inducible CLTM®)
- SGM: until the positive is readable

Antibiotic Susceptibility test (AST)

Plate Code:		RAPMYCO												
	1	2	3	4	5	6	7	8	9	10	11	12	ANTIMICROBICS	
A	SXT 0.25/4.75	SXT 0.5/9.5	SXT 1/19	SXT 2/38	SXT 4/76	SXT 8/152	LZD 1	LZD 2	LZD 4	LZD 8	LZD 16	LZD 32	SXT	Trimethoprim / sulfamethoxazole
B	CIP 0.12	CIP 0.25	CIP 0.5	CIP 1	CIP 2	CIP 4	IMI 2	IMI 4	IMI 8	IMI 16	IMI 32	IMI 64	CIP	Ciprofloxacin
C	MXF 0.25	MXF 0.5	MXF 1	MXF 2	MXF 4	MXF 8	FEP 1	FEP 2	FEP 4	FEP 8	FEP 16	FEP 32	MXF	Moxifloxacin
D	FOX 4	FOX 8	FOX 16	FOX 32	FOX 64	FOX 128	AUG2 2/1	AUG2 4/2	AUG2 8/4	AUG2 16/8	AUG2 32/16	AUG2 64/32	FOX	Cefoxitin
E	AMI 1	AMI 2	AMI 4	AMI 8	AMI 16	AMI 32	AMI 64	AXO 4	AXO 8	AXO 16	AXO 32	AXO 64	AMI	Amikacin
F	DOX 0.12	DOX 0.25	DOX 0.5	DOX 1	DOX 2	DOX 4	DOX 8	DOX 16	MIN 1	MIN 2	MIN 4	MIN 8	DOX	Doxycycline
G	TGC 0.015	TGC 0.03	TGC 0.06	TGC 0.12	TGC 0.25	TGC 0.5	TGC 1	TGC 2	TGC 4	TOB 1	TOB 2	TOB 4	TGC	Tigecycline
H	CLA 0.06	CLA 0.12	CLA 0.25	CLA 0.5	CLA 1	CLA 2	CLA 4	CLA 8	CLA 16	TOB 8	TOB 16	POS	CLA	Clarithromycin
													LZD	Linezolid
													IMI	Imipenem
													FEP	Cefepime
													AUG2	Amoxicillin / clavulanic acid 2:1 ratio
													AXO	Ceftriaxone
													MIN	Minocycline
													TOB	Tobramycin
													POS	Positive Control

Antibiotic Susceptibility test (AST)

Plate Code: **SLOMYCOI**

	1	2	3	4	5	6	7	8	9	10	11	12
A	CLA 0.06	CLA 0.12	CLA 0.25	CLA 0.5	CLA 1	CLA 2	CLA 4	CLA 8	CIP 16	STR 64	DOX 16	ETH 20
B	CLA 16	CLA 32	CLA 64	MXF 8	RIF 8	SXT 8/152	AMI 64	LZD 64	CIP 8	STR 32	DOX 8	ETH 10
C	RFB 8	EMB 16	INH 8	MXF 4	RIF 4	SXT 4/76	AMI 32	LZD 32	CIP 4	STR 16	DOX 4	ETH 5
D	RFB 4	EMB 8	INH 4	MXF 2	RIF 2	SXT 2/38	AMI 16	LZD 16	CIP 2	STR 8	DOX 2	ETH 2.5
E	RFB 2	EMB 4	INH 2	MXF 1	RIF 1	SXT 1/19	AMI 8	LZD 8	CIP 1	STR 4	DOX 1	ETH 1.2
F	RFB 1	EMB 2	INH 1	MXF 0.5	RIF 0.5	SXT 0.5/9.5	AMI 4	LZD 4	CIP 0.5	STR 2	DOX 0.5	ETH 0.6
G	RFB 0.5	EMB 1	INH 0.5	MXF 0.25	RIF 0.25	SXT 0.25/4.75	AMI 2	LZD 2	CIP 0.25	STR 1	DOX 0.25	ETH 0.3
H	RFB 0.25	EMB 0.5	INH 0.25	MXF 0.12	RIF 0.12	SXT 0.12/2.38	AMI 1	LZD 1	CIP 0.12	STR 0.5	DOX 0.12	POS

ANTIMICROBICS	
CLA	Clarithromycin
RFB	Rifabutin
EMB	Ethambutol
INH	Isoniazid
MXF	Moxifloxacin
RIF	Rifampin
SXT	Trimethoprim / sulfamethoxazole
AMI	Amikacin
LZD	Linezolid
CIP	Ciprofloxacin
STR	Streptomycin
DOX	Doxycycline
ETH	Ethionamide
POS	Positive Control

Antibiotic Susceptibility test (AST) Interpretation

CLINICAL AND LABORATORY STANDARDS INSTITUTE®
3rd Edition

M24

Susceptibility Testing of Mycobacteria, *Nocardia* spp., and Other Aerobic Actinomycetes

CLINICAL AND LABORATORY STANDARDS INSTITUTE®
1st Edition

M62

Performance Standards for Susceptibility Testing of Mycobacteria, *Nocardia* spp., and Other Aerobic Actinomycetes

			Sensitive	Intermediate	Resistant
Etude <i>in vitro</i> de la sensibilité aux antibiotiques par la technique Sensititre® (broth microdilution method for rapidly growing mycobacteria) - = Sensititre RAPMYCO	Linezolid	µg/ml	≤8	16	≥32
	Clarithromycine	µg/ml	≤2	4	≥8
	Amikacine	µg/ml	≤16	32	≥64
	Cefoxitine	µg/ml	≤16	32-64	≥128
	Ceftriaxone	µg/ml	NO CLSI value		
	Imipenem	µg/ml	≤4	8-16	≥32
	Tobramycine	µg/ml	≤2	4	≥8
	Ciprofloxacine	µg/ml	≤1	2	≥4
	Moxifloxacine	µg/ml	≤1	2	≥4
	Minocycline	µg/ml	NO CLSI value		
	Amoxicilline/acide Clavulanique	µg/ml	NO CLSI value		
	Triméthoprime/sulfaméthoxazole	µg/ml	≤2/38	/	≥4/76
	Cefepime	µg/ml	No CLSI value		
Tigecycline	µg/ml	No CLSI value			
Doxycycline	µg/ml	≤1	2-4	≥8	

Etude <i>in vitro</i> de la sensibilité aux antibiotiques par la technique Sensititre® (broth microdilution method for slowly growing mycobacteria) - = Sensititre SLOMYCO kansasii	Clarithromycine	µg/ml	≤8	16	≥32
	Rifampicine	µg/ml	≤1	/	≥2
	Amikacine	µg/ml	≤16	32	≥64
	Ciprofloxacine	µg/ml	≤1	2	≥4
	Ethambutol	µg/ml	No CLSI value		
	Linezolid	µg/ml	≤8	16	≥32
	Moxifloxacine	µg/ml	≤1	2	≥4
	Rifabutine	µg/ml	≤2	/	≥4
	Triméthoprime/sulfaméthoxazole	µg/ml	≤2/38	/	≥4/76
	Doxycycline	µg/ml	≤1	2-4	≥8
	Isoniazide	µg/ml	No CLSI value		
	Streptomycine	µg/ml	No CLSI value		
	Ethionamide	µg/ml	No CLSI value		

Etude <i>in vitro</i> de la sensibilité aux antibiotiques par la technique Sensititre® (broth microdilution method for slowly growing mycobacteria) - = Sensititre SLOMYCO MAC (<i>Mycobacterium avium</i> complex)	Clarithromycine	µg/ml	≤8	16	≥32
	Rifampicine	µg/ml	No CLSI value		
	Amikacine (IV)	µg/ml	≤16	32	≥64
	Amikacine (liposomal inhalé)	µg/ml	≤64	-	≥128
	Ciprofloxacine	µg/ml	No CLSI value		
	Ethambutol	µg/ml	No CLSI value		
	Linezolid	µg/ml	≤8	16	≥32
	Moxifloxacine	µg/ml	≤1	2	≥4
	Rifabutine	µg/ml	No CLSI value		
	Triméthoprime/sulfaméthoxazole	µg/ml	No CLSI value		
	Doxycycline	µg/ml	No CLSI value		
	Isoniazide	µg/ml	No CLSI value		
	Streptomycine	µg/ml	No CLSI value		
Ethionamide	µg/ml	No CLSI value			

Etude <i>in vitro</i> de la sensibilité aux antibiotiques par la technique Sensititre® (broth microdilution method for slowly growing mycobacteria) - = Sensititre SLOMYCO other than MAC and kansasii	Clarithromycine	µg/ml	≤8	16	≥32
	Rifampicine	µg/ml	≤1	/	≥2
	Amikacine	µg/ml	≤16	32	≥64
	Ciprofloxacine	µg/ml	≤1	2	≥4
	Ethambutol	µg/ml	No CLSI value		
	Linezolid	µg/ml	≤8	16	≥32
	Moxifloxacine	µg/ml	≤1	2	≥4
	Rifabutine	µg/ml	≤2	/	≥4
	Triméthoprime/sulfaméthoxazole	µg/ml	≤2/38	/	≥4/76
	Doxycycline	µg/ml	≤1	2-4	≥8
	Isoniazide	µg/ml	No CLSI value		
	Streptomycine	µg/ml	No CLSI value		
	Ethionamide	µg/ml	No CLSI value		

Epidemiology



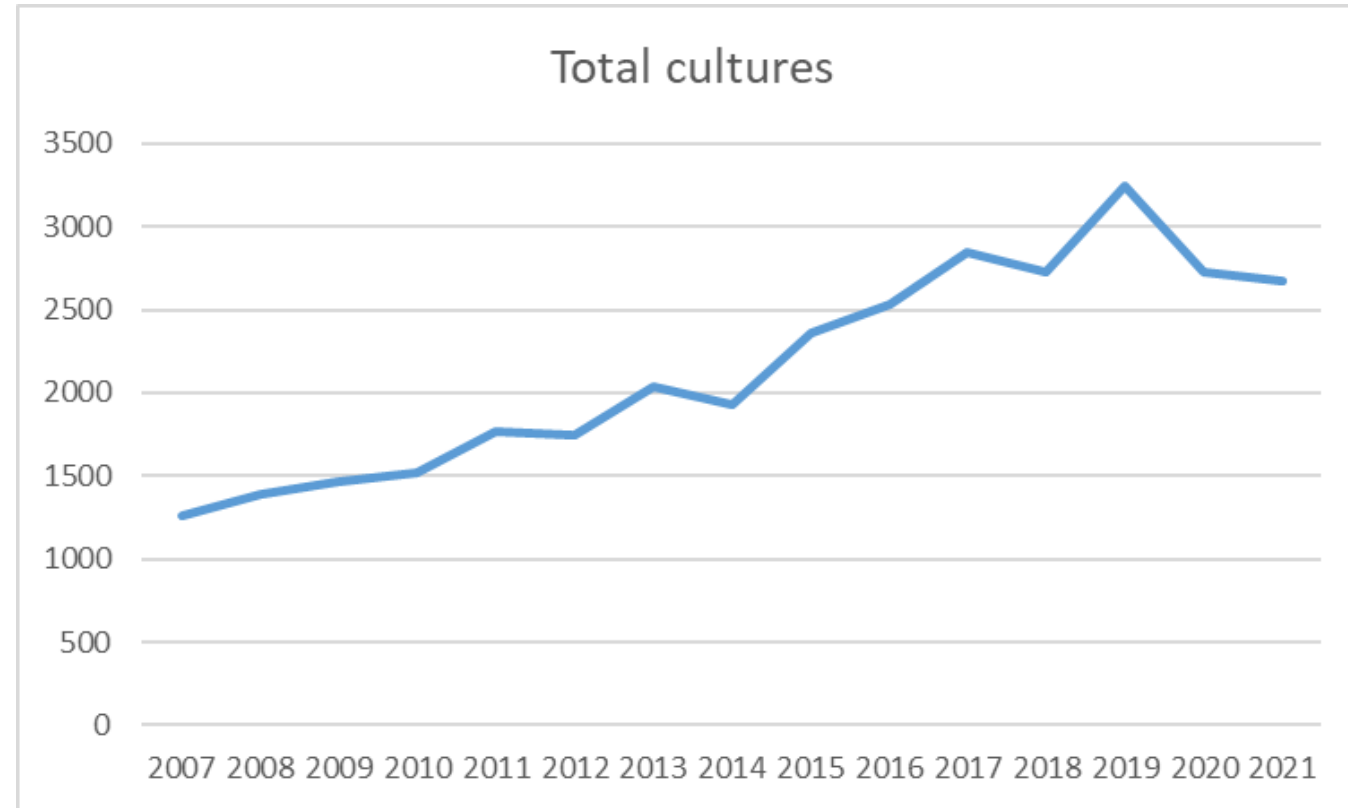
Principal laboratory performing AST for clinically relevant cases



NTM surveillance/diagnosis data from the Belgian NRC since 2007

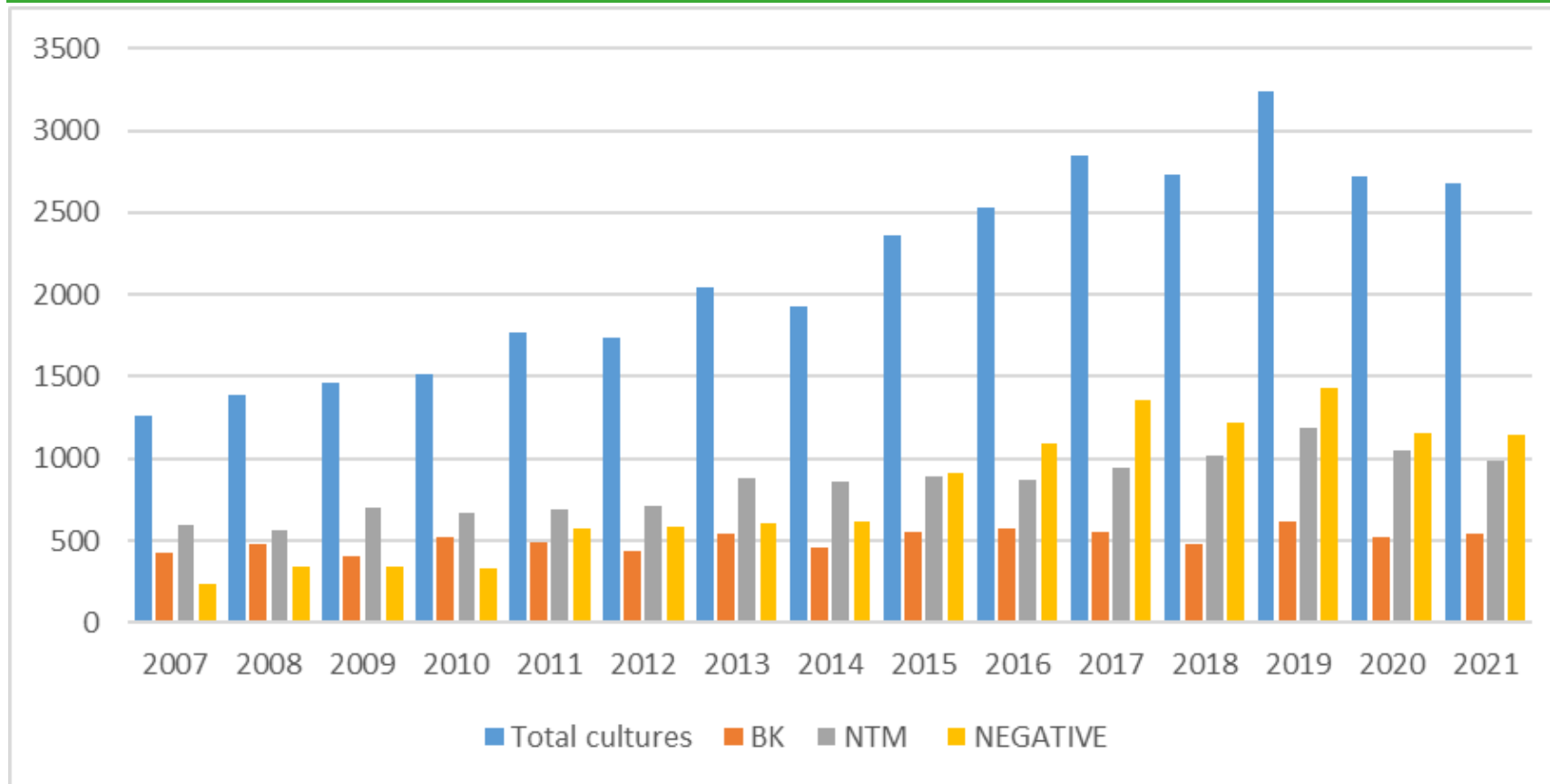
Evolution of the laboratory analysis

Year	Total cultures
2007	1265
2008	1390
2009	1462
2010	1517
2011	1765
2012	1743
2013	2042
2014	1929
2015	2366
2016	2534
2017	2851
2018	2729
2019	3243
2020	2725
2021	2674



Soetaert Karine, Subissi Lorenzo, Ceysens Pieter-Jan, Vanfleteren Brigitte, Chantrenne Marianne, Asikainen Tommi, Duysburgh Els, Mathys Vanessa. Strong increase of true and false positive mycobacterial cultures sent to the National Reference Centre in Belgium, 2007 to 2016. Euro Surveill. 2019;24(11):pii=1800205. <https://doi.org/10.2807/1560-7917.ES.2019.24.11.1800205>

Evolution of the laboratory analysis



Increasing numbers of NTM



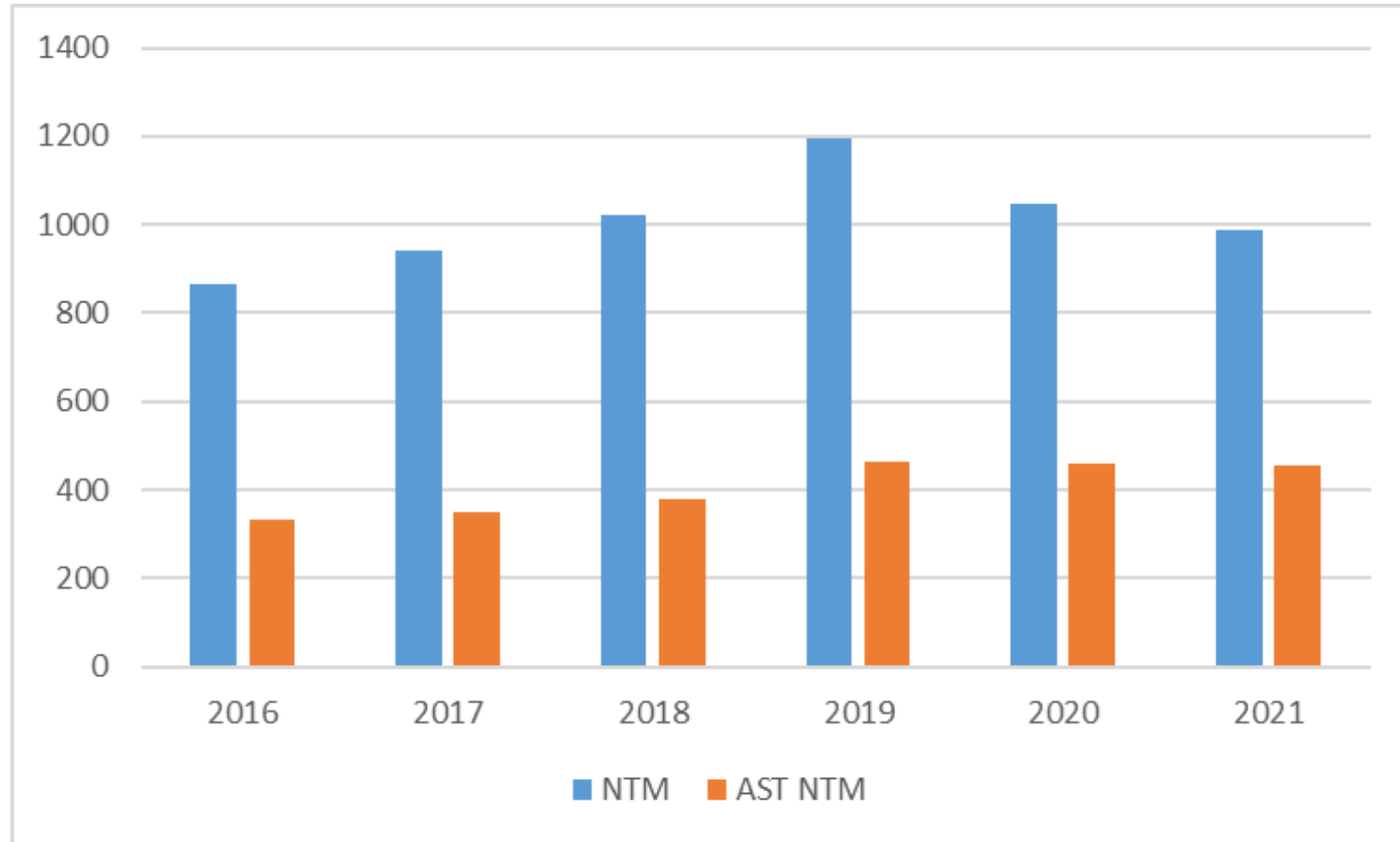
- improved culturing technique
- increased awareness
- increase in disease incidence

Soetaert Karine, Subissi Lorenzo, Ceyssens Pieter-Jan, Vanfleteren Brigitte, Chantrenne Marianne, Asikainen Tommi, Duysburgh Els, Mathys Vanessa. Strong increase of true and false positive mycobacterial cultures sent to the National Reference Centre in Belgium, 2007 to 2016. Euro Surveill. 2019;24(11):pii=1800205.

<https://doi.org/10.2807/1560-7917.ES.2019.24.11.1800205>

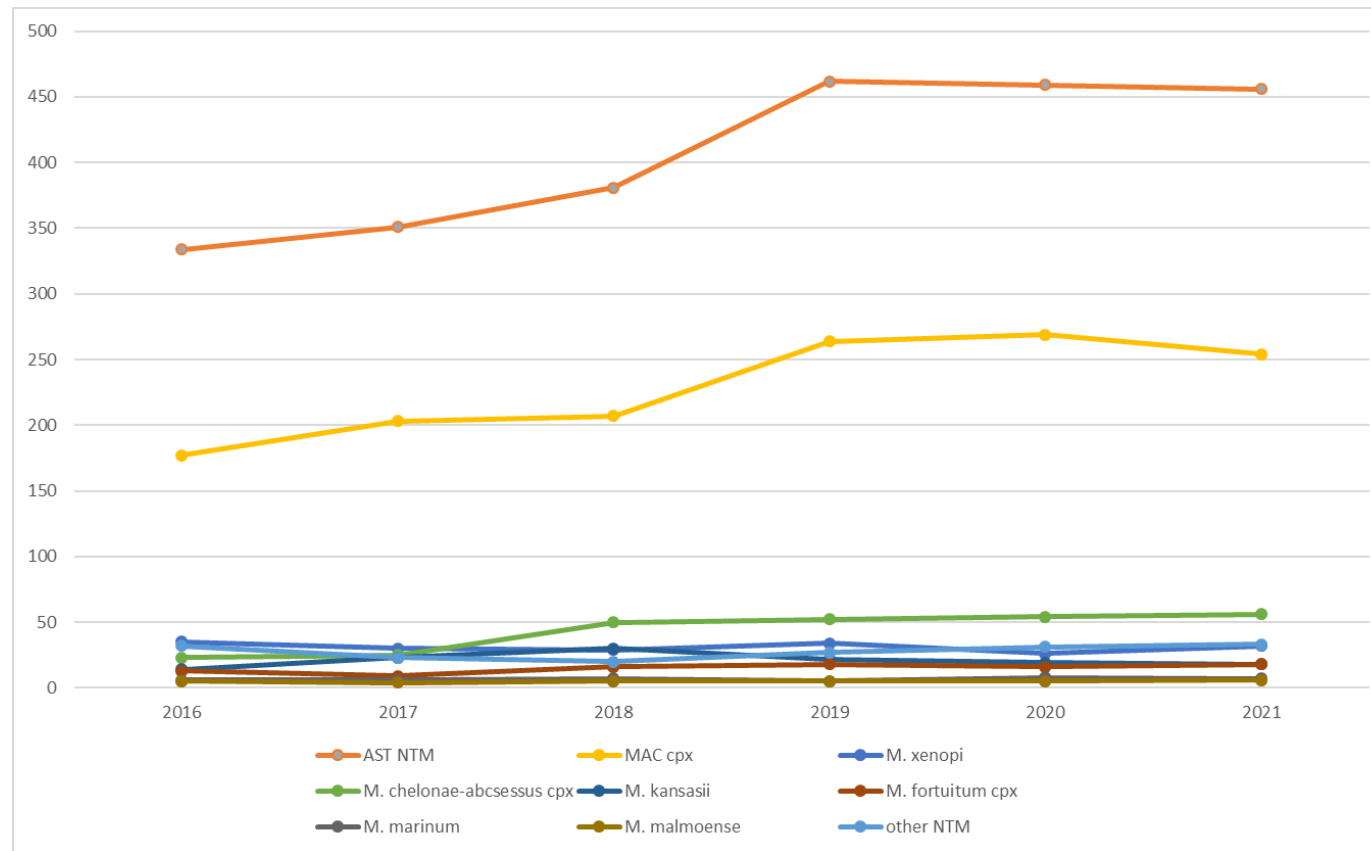
Evolution of the laboratory analysis

Year	NTM	AST NTM
2016	867	334
2017	941	351
2018	1022	381
2019	1193	462
2020	1046	459
2021	988	456



AST on NTM species is only performed upon physicians' request.

Evolution of the laboratory analysis



→ The correct identification of the aetiological agent is critical for diagnosis and patient management, as species differ in clinical relevance, antibiotic susceptibility and treatment outcome

Thank you for your attention

NRC Mycobacterium

Vanessa Mathys

Karine Soetaert

Samira Boarbi

Brigitte Vanfleteren

Mehdi Kiass

Romuald Lizon

Pierre-Yves Adnet

Tatiana Theeten

Kevin Charlier

Laboratory of Medical Microbiology

Alexandra Vodolazkaia

Marina Mukovnikova

